ALTERNATIVE CONSTRUCTION & ENVIRONMENTAL SOLUTIONS, INC.

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Augusta, GA 30904

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January 23, 2024

Client: Augusta-Richmond County Central Services Department
2760 Peach Orchard Road
Augusta, GA 30906
Attn: Ms. Maria Rivera-Rivera
mrivera-rivera@augustaga.gov

Project: Indoor Air Quality Inspection and Spore Count Analysis
The Boathouse Community Center
101 Riverfront Drive
Augusta, GA
Report Number: 2006-113-002

1.0 SCOPE

At the request of the client, a representative of Alternative Construction & Environmental Solutions, Inc., (ACES) conducted an Indoor Air Quality visual inspection, mold sampling and analysis of the Boathouse Community Center at 101 Riverfront Drive in Augusta, Georgia. The areas tested were limited to be representative of the climate-controlled portion of the building. Sampling was not conducted in every space of the building.

2.0 SITE/VISUAL INSPECTION

A representative of ACES conducted a visual site inspection of the above-referenced project area on January 19, 2024. This was performed to determine the current condition of the surfaces and to identify any discoloration or staining at the time of this inspection. Visible staining indicative of mold colonization was observed on ceiling tiles, walls and insulation backing in the Kitchen. No destructive or invasive inspecting was performed. A visual inspection performed in the basement area occupied by the Rowing Club identified staining indicative of mold colonization on the drywall ceilings. Air sampling was not conducted in the basement as this area was not under climate control.

3.0 DISCUSSION

Sampling for mold is typically to aid in establishing 1) whether mold spores are present in an area that was designed to prevent mold from entering or forming, 2) if there is an inside source of spores

that could lead to increased occupants' exposure, or 3) if there is moisture and the presence of fungi that could be an indication of possible structural damage now or sometime in the future.

- ***Air Samples:***
  Air samples are divided into two different methods, culturable and non-culturable (non-viable or "spore trap"). Both samples are acquired using an air pump attached to a media.

  A. BioCassettes® with a Malt Extract Agar (MEA) are used for the determination of culturable (viable) fungi present in the air. Air sample duration is from one to five minutes at 28.3 liters per minute based on visual evaluation of the areas level of suspected contamination.

  B. Non-culturable samples are acquired using Air-O-Cell® spore trap samplers. This is a particulate sampling cassette for the rapid collection and analysis of a wide range of mold spores. This type of sampling does not allow differentiation between viable and non-viable mold spores. Air sample duration is determined by an expected level of contamination chart with sample times of 0.5 – 10 minutes at 15 liters per minute.

  "Air Sampling is limited, and negative results do not document the absence of mold exposure. For example, mold may be growing in carpets or on walls and wallpapers, yet not be airborne at the time of sampling. Where there are other indications, such as moisture noted where it should not be, further investigation for hidden sources is indicated." (ref. 1)

- ***Surface Samples:***
  Surface samples are typically taken by tape lift imprint, by swabbing the surface of suspected mold growth with a culturette swab, or by destructive sampling of the suspected mold growth materials. These samples are submitted to the laboratory for analysis for microbial contamination. The laboratory reports the findings by direct microscopic examination to identify the types of mold growth.

"There is substantial natural variability in the amount of mold in air. Understandably, the EPA and other government agencies have not set numeric standards for indoor concentrations of mold or mold spores." (ref. 1)

4.0 SAMPLING AND ANALYSIS

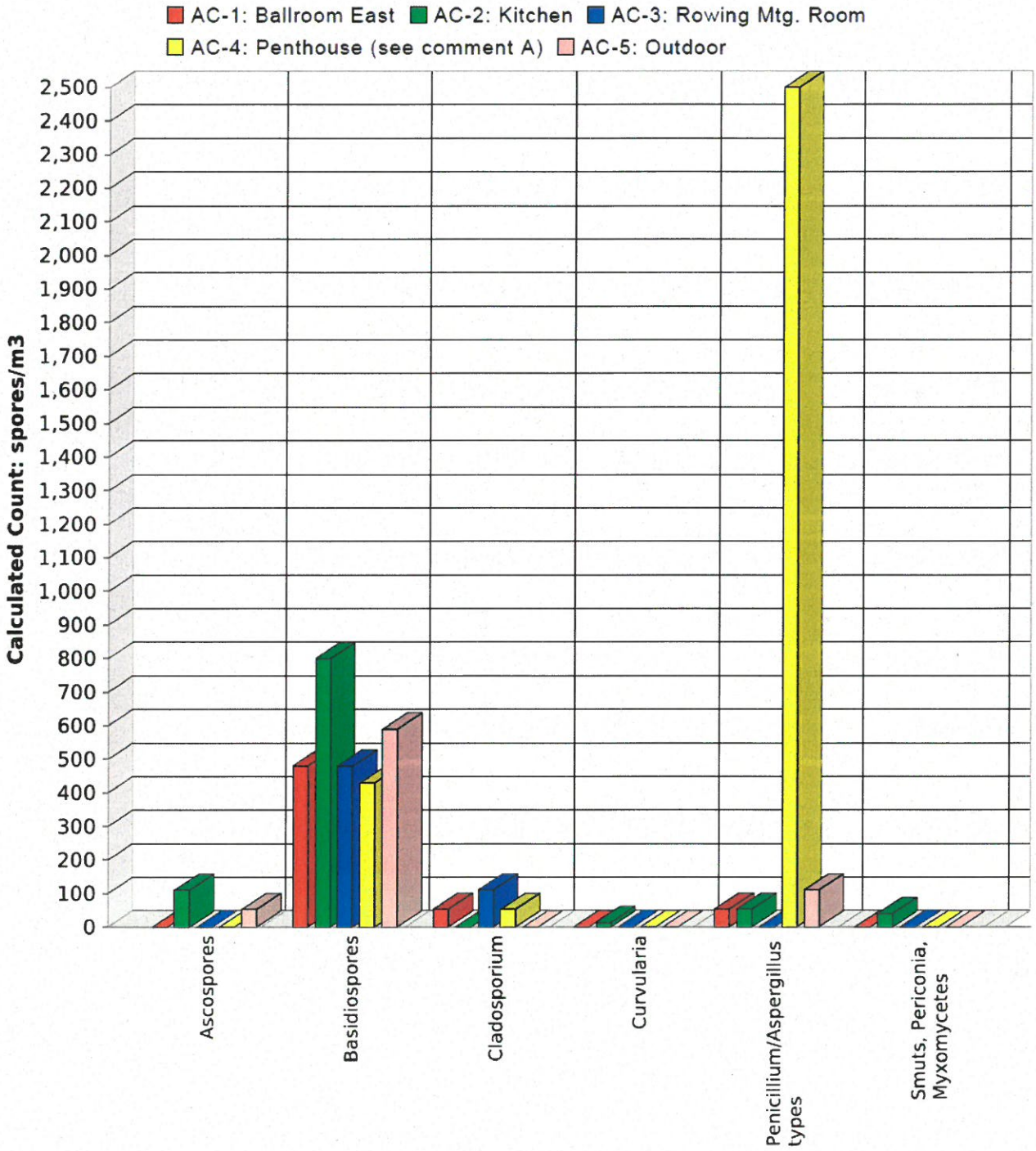
Sampling was accomplished utilizing a Zefon Bio-Pump with Air-O-Cell® cassettes. Five Air-O-Cell® samples were taken from various areas to include both inside and outside samples. The number of samples taken during this investigation was specified by the client.

Samples collected were assigned a unique sample ID number and placed in a sealed container. Samples were sent to EMLab P&K (Environmental Microbiology Laboratory, Inc.), an American Industrial Hygiene Association (AIHA) Environmental Microbiology-accredited laboratory (#178699).

Table I presents a graphical depiction of the spore counts obtained from this project. Sample AC-1 is depicted in red for the inside sample obtained from the Ballroom. Sample AC-2 is depicted in green

for the inside sample obtained from the Kitchen. Sample AC-3 is depicted in blue for the inside sample obtained from the Rowing Meeting Room. Sample AC-4 is depicted in yellow for the inside sample obtained from the Penthouse. Sample AC-5 is depicted in pink and represents the outdoor sample.

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY



Comment A: 41 of the raw count Penicillium/Aspergillus type spores were present as a single clump.

The Bioaerosol Committee of the American Conference of Governmental Industrial Hygienists (ACGIH) states that outdoor airborne fungi concentration "routinely exceeds 1,000 CFU/m^3 and may average near 10,000 CFU/m^3 in summer months." No occupational exposure limit for bioaerosols has been promulgated by the Occupational Safety and Health Administration (OSHA). (ref. 3)

Additionally, temperature, relative humidity and carbon monoxide readings were taken using the Gray Wolf Indoor Air Quality Probe MP Surveyor Pro with serial number 78014.

Location	Time	Temp. (°F)	Relative Humidity (%)	Carbon Monoxide (ppm)
Ballroom	1010	58.7	35.2	1.0
Kitchen	1017	57.9	37.6	0.4
Rowing Meeting Room	1027	53.2	44.1	0.3
Penthouse	1038	73.0	41.1	0.3
Outdoor	1055	51.7	54.8	0.4

Elevated Carbon Monoxide levels in indoor air indicate the existence of a combustion source exhaust that is not effectively being ventilated out of the building. The standard of 25 ppm established by the American Conference of Governmental Industrial Hygienist (ACGIH) is widely accepted as a limit for defining hazardous carbon monoxide levels. (ref. 3) The ASHRAE ventilation standard has established 9 ppm or greater of carbon monoxide within an occupied space as being a level of concern. (ref. 2) The Environmental Protection Agency defines 9 ppm as a limit for acceptable air quality.

"The Relative Humidity (RH) of the indoor air and ventilation system should be below 60 percent. Ideally, RH should be kept between 30 and 50 percent because, at a RH of 50 percent or more, hydroscopic dust will absorb water that may allow the growth of fungi and house dust mites on indoor surfaces." (ref. 1)

5.0 ATTACHMENTS

Attachment I Lab Results
Attachment II Photographs

6.0 CONCLUSIONS

There was visible staining indicative of mold growth found on visible surfaces inside the project area. These included ceiling tiles, walls and insulation backing in the kitchen. Although spore count samples were not collected from the basement (Rowing Club facility), visual staining indicative of mold colonization was observed and photographed on the ceilings throughout. No destructive or invasive inspecting was performed.

Spores of Cladosporium, Curvularia and Smuts/Periconia/Myxomycetes were identified in the inside samples but were not identified in the outside sample. Basidiospores and, particularly, Penicillium/Aspergillus type spores were identified inside in significantly higher concentrations than

in the outdoor sample. This data does suggest indoor sources for these spore types at the time of this inspection.

The following is a list of the spores found inside the project area with a brief description of each.

Ascospores are found everywhere in nature, particularly in Saprophytes and plant pathogens. The spores are predominantly forcibly discharged during periods of high humidity or rain.

Basidiospores is a common spore found outdoors on decaying wood and on plants. In the indoor environment, it is one of the fungi responsible for causing white or brown wood rot if allowed to form large colonies and can sometimes be recognized by the presence of mushrooms. These spores can grow to destroy the structural wood of buildings. Known health effects in immunocompromised individuals can include type 1 allergies.

Cladosporium is typically found on leaves and decaying plants in the outdoor environment. In the indoor environment, it can be found in insulation and on window panes that have colder surfaces.

Curvularia in the indoor environment is typically from plant debris, soil, and facultative plant pathogens of tropical or subtropical plants. More commonly found in tropical, subtropical regions.

Penicillium/Aspergillus in the outdoor environment is often found in soil, decaying plants and in stored grain. Indoors it is often found on water-damaged building materials such as wallboard, chipboards and decaying fabrics or foodstuffs like cheese, herbs and onions. Known health effects in immunocompromised individuals can include hay fever, asthma and type 3 hypersensitivity pneumonitis.

According to the MoldRANGE™ Extended Outdoor Comparison Chart, some of the spore counts recorded in this survey were above the typical very high outdoor data comparison for the month of January in the State of Georgia.

No occupational exposure limit for bioaerosols has been promulgated by the Occupational Safety and Health Administration (OSHA). "If fungal concentrations indoors are consistently higher than those outdoors, then indoor sources are indicated." (ref. 3)

Most spores found during this sampling are primarily found in the soil, leaves, grass, weeds, and other types of vegetation and can be easily transmitted into the building by people moving in and out.

The Relative Humidity levels found inside during this inspection were below 50%. A Relative Humidity of 50% or greater can increase the chances of fungi growth. It should be noted that not all of the HVAC units were operational at the time of sampling.

If any materials are found in the future with visual mold growth or damaged appearance, they should be cleaned or removed/discarded as per EPA guidelines published in "Mold Remediation in Schools and Commercial Buildings" (EPA 402-K-01-001). (ref. 4)

It should be noted that microbiological growth can and probably will reoccur if any source of moisture is not corrected and maintained as appropriate. Many spores are naturally-occurring and will grow again with proper moisture, temperature and food source.

This report relates only for this time and conditions present during our investigation. Facilities are constantly under influence from several external factors, activities, environmental conditions and pollutants that are subject to change. If any new or conflicting information becomes available at a later date, please advise ACES and any appropriate revisions and/or comments will be made. This report was prepared for the client and should not be reproduced, except in whole and only with the written approval of ACES.

ALTERNATIVE CONSTRUCTION & ENVIRONMENTAL SOLUTIONS, INC.

Dan D. Troutman
President

Steve Connor
Project Manager

Attachment

References:

1. University of Connecticut Health Center Division of Occupational and Environmental Medicine, Center for Indoor Environments and Health, *Guidance for Clinicians on the Recognition and Management of Health Effects Related to Mold Exposure and Moisture Indoors, (September 30, 2004)*

2. American Society of Heating, Refrigerating and Air-Conditioning Engineers, Inc. (*ANSI/ASHRAE 62-1989), Ventilation for Acceptable Air Quality*, ASHRAE, Atlanta, GA 1992.

3. American Conference of Governmental Industrial Hygienists (ACGIH), *Bioaerosols: Assessment and Control* (Edited by J. Marcher), Cincinnati, OH 1999.

4. Environmental Protection Agency (EPA), *Mold Remediation in Schools and Commercial Buildings (*EPA 402-K-01-001, March 2001)

eurofins | Built Environment Testing

Report for:

Mr. Steve Connor
Alternative Construction and Environmental Solutions
P.O. Box 3229
Augusta, GA 30914

Regarding: Eurofins EPK Built Environment Testing, LLC
Project: 2006-113-002; Boathouse
EML ID: 3512408

Approved by:

Business Unit Manager
Balu Krishnan

Dates of Analysis:
Spore trap analysis: 01-23-2024

Service SOPs: Spore trap analysis (EB-MY-S-1038)
AIHA-LAP, LLC accredited service, Lab ID #221504

All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank correction of results is not applied. The results relate only to the samples as received and tested. Information supplied by the client which can affect the validity of results: sample air volume.

Eurofins EPK Built Environment Testing, LLC ("the Company"), a member of the Eurofins Built Environment Testing group of companies, shall have no liability to the client or the client's customer with respect to decisions or recommendations made, actions taken or courses of conduct implemented by either the client or the client's customer as a result of or based upon the Test Results. In no event shall the Company be liable to the client with respect to the Test Results except for the Company's own willful misconduct or gross negligence nor shall the Company be liable for incidental or consequential damages or lost profits or revenues to the fullest extent such liability may be disclaimed by law, even if the Company has been advised of the possibility of such damages, lost profits or lost revenues. In no event shall the Company's liability with respect to the Test Results exceed the amount paid to the Company by the client therefor.
Eurofins EPK Built Environment Testing, LLC's LabServe® reporting system includes automated fail-safes to ensure that all AIHA-LAP, LLC quality requirements are met and notifications are added to reports when any quality steps remain pending.

Eurofins EPK Built Environment Testing, LLC
6215 Regency Parkway, Suite 900, Norcross, GA 30071
(866) 871-1984 www.eurofinsus.com/Built

Client: Alternative Construction and Environmental Solutions
C/O: Mr. Steve Connor
Re: 2006-113-002; Boathouse

Date of Sampling: 01-19-2024
Date of Receipt: 01-22-2024
Date of Report: 01-23-2024

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	AC-1: Ballroom East		AC-2: Kitchen		AC-3: Rowing Mtg. Room		AC-4: Penthouse		AC-5: Outdoor	
Comments (see below)	None		None		None		A		None	
Lab ID-Version‡:	17150027-1		17150028-1		17150029-1		17150030-1		17150031-1	
Analysis Date:	01/23/2024		01/23/2024		01/23/2024		01/23/2024		01/23/2024	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria										
Arthrinium										
Ascospores			2	110					1	53
Aureobasidium										
Basidiospores	9	480	15	800	9	480	8	430	11	590
Bipolaris/Drechslera group										
Botrytis										
Chaetomium										
Cladosporium	1	53			2	110	1	53		
Curvularia			1	13						
Epicoccum										
Fusarium										
Myrothecium										
Nigrospora										
Other colorless										
Penicillium/Aspergillus types†	1	53	1	53			77	2,500	2	110
Pithomyces										
Rusts										
Smuts, Periconia, Myxomycetes			3	40						
Stachybotrys										
Stemphylium										
Torula										
Ulocladium										
Zygomycetes										
Background debris (1-4+)	2+		2+		1+		2+		2+	
Sample volume (liters)	75		75		75		75		75	
§ TOTAL SPORES/m3		590		1,000		590		2,900		750

Comments: A) 41 of the raw count *Penicillium/Aspergillus* type spores were present as a single clump.

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.
† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium, Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.
††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m^3 divided by the raw count, expressed in spores/m^3, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.
‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".
§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.

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Re: 2006-113-002; Boathouse

Date of Sampling: 01-19-2024
Date of Receipt: 01-22-2024
Date of Report: 01-23-2024

MoldRANGE™, Local Climate; Extended Outdoor Comparison

Outdoor Location: AC-5, Outdoor

Fungi Identified	Outdoor data	Typical Outdoor Data for: January in Southeast† EMLab Regional Climate code¹ A Annual Temp, A Elev., B Rain, A Temp. Range (n‡=264)						Typical Outdoor Data for: The entire year in Southeast† EMLab Regional Climate code¹ A Annual Temp, A Elev., B Rain, A Temp. Range (n‡=3474)					
Project zip code 30901	spores/m3	very low	low	med	high	very high	freq %	very low	low	med	high	very high	freq %
Generally able to grow indoors*													
Alternaria	-	12	13	27	32	53	16	13	13	27	80	120	40
Bipolaris/Drechslera group	-	-	-	-	-	-	6	13	13	20	53	93	21
Chaetomium	-	-	-	-	-	-	3	13	13	13	40	85	5
Cladosporium	-	27	53	160	480	1,000	80	53	130	600	1,800	3,200	91
Curvularia	-	11	13	13	35	65	9	13	13	27	80	150	31
Nigrospora	-	13	13	13	22	27	10	7	13	13	40	53	20
Penicillium/Aspergillus types	110	25	40	120	320	630	73	44	67	210	670	1,200	75
Stachybotrys	-	-	-	-	-	-	1	10	13	27	80	190	1
Torula	-	-	-	-	-	-	4	7	13	13	44	77	12
Seldom found growing indoors**													
Ascospores	53	13	34	110	430	750	70	53	110	530	1,900	3,500	90
Basidiospores	590	40	93	800	3,000	7,200	92	160	400	2,300	11,000	22,000	98
Rusts	-	-	-	-	-	-	3	13	13	27	67	150	19
Smuts, Periconia, Myxomycetes	-	13	13	27	53	82	55	13	20	53	150	230	71
§ TOTAL SPORES/m3	750												

¹EMLab Regional Climate codes are a climate classification scheme for regional geographic areas containing multiple states. The MoldRANGE™ Local Climate report uses the sampling location zip code to identify the EMLab Regional Climate code in that area. Using information available from the NOAA weather database, the EMLab Regional Climate code sharpens the precision of the MoldRANGE™reporting system, providing more reliable estimates of the range and average concentrations of the different airborne fungal spore types for each region. Additional information on the EMLab Regional Climate code system can be found on the last page of this report.

†The Typical Outdoor Data represents the typical outdoor spore levels across the region's group of states for the time period and EMLab Regional Climate code indicated. The last column represents the frequency of occurrence. The very low, low, med, high, and very high values represent the 10, 20, 50, 80, and 90 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 20% of the time it is present in levels above the detection limit and below 53 spores/m3. These values are updated periodically and if not enough data is available to make a statistically meaningful assessment, it is indicated with a dash.

‡ n is the sample size used to calculate the MoldRANGE™ Local Climate data summarized in the table.

* The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

** These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.

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Understanding EMLab Regional Climate Codes

Outdoor airborne spore concentrations are strongly influenced by climate and weather patterns, often resulting in pronounced seasonal and diurnal cycles (Burge 1995). The seasonal climatic changes directly affect the growth cycle of plants, thereby influencing fungal growth, spore maturation, and release cycles. By evaluating outdoor spore concentrations across similar climatic zones rather than for the state as a whole, it is possible to provide a more representative estimate of typical outdoor spore levels and frequency of occurrence for different airborne fungal spore types in a given area.

The EMLab Regional Climate code system is a novel classification system that uses data from the NOAA - National Oceanic and Atmospheric Administration database to define unique climate zones. The following climate variables, for each regional zip code, are obtained from NOAA and assigned a letter code of A (above the regional average for that variable) or B (below the regional average for that variable):

1. Annual High Temperature
2. Elevation
3. Rainfall/Precipitation
4. Monthly Temperature Range

The result is a 4-character code assigned to each statewide zip code, referred to as the Regional Climate Code. Below are some examples of decoded Regional Climate Codes:

AAAA = Above avg. Annual High Temperature, Above avg. Elevation, Above avg. Rainfall/Precipitation, Above avg. Monthly Temperature Range
AABB = Above avg. Annual High Temperature, Above avg. Elevation, Below avg. Rainfall/Precipitation, Below avg. Monthly Temperature Range
BBAA = Below avg. Annual High Temperature, Below avg. Elevation, Above avg. Rainfall/Precipitation, Above avg. Monthly Temperature Range

The actual outdoor air sample data from matching regional climate codes in each group of states are then compiled in a manner relating typical spore concentrations and frequency of occurrence.

The data presented in this report is from the Southeast Region which includes the states of: AL, FL, GA, NC, SC, and VA

The NOAA regional climate variables were selected by mapping data points from a subset of approximately 145,000 weather and geographic database entries to over 80,000 outdoor spore trap samples with known zip codes and assessing them using orthogonal array experimental design techniques. The results were then compared to the typical ranges of spore types found when grouping zip codes using the Koppen-Geiger climatic classification system; a commonly used climatic system that provides an objective numerical definition in terms of climatic elements such as temperature, rainfall, and other seasonal characteristics . The EMLab Regional Climate codes showed improved granularity and refinement of the zip code groupings, implying a better representation of the expected range of spore types to be found within an individual zip code.

The values on this report were calculated by obtaining the four variables listed above from the over 585 million data points of weather and geographic information available in the NOAA database, and determining the frequencies and percentile values of spore types by utilizing over 180,000 Eurofins EMLab P&K outdoor spore trap samples with known zip codes.

This report groups regional zip codes in relation to these EMLab Regional Climate codes and summarizes MoldRANGE™ data by month and year within each EMLab Regional Climate code.

References:

Burge, Harriet, A. Bioaerosols: Boca Raton: Lewis Publishers, pp. 163-171, 1995.

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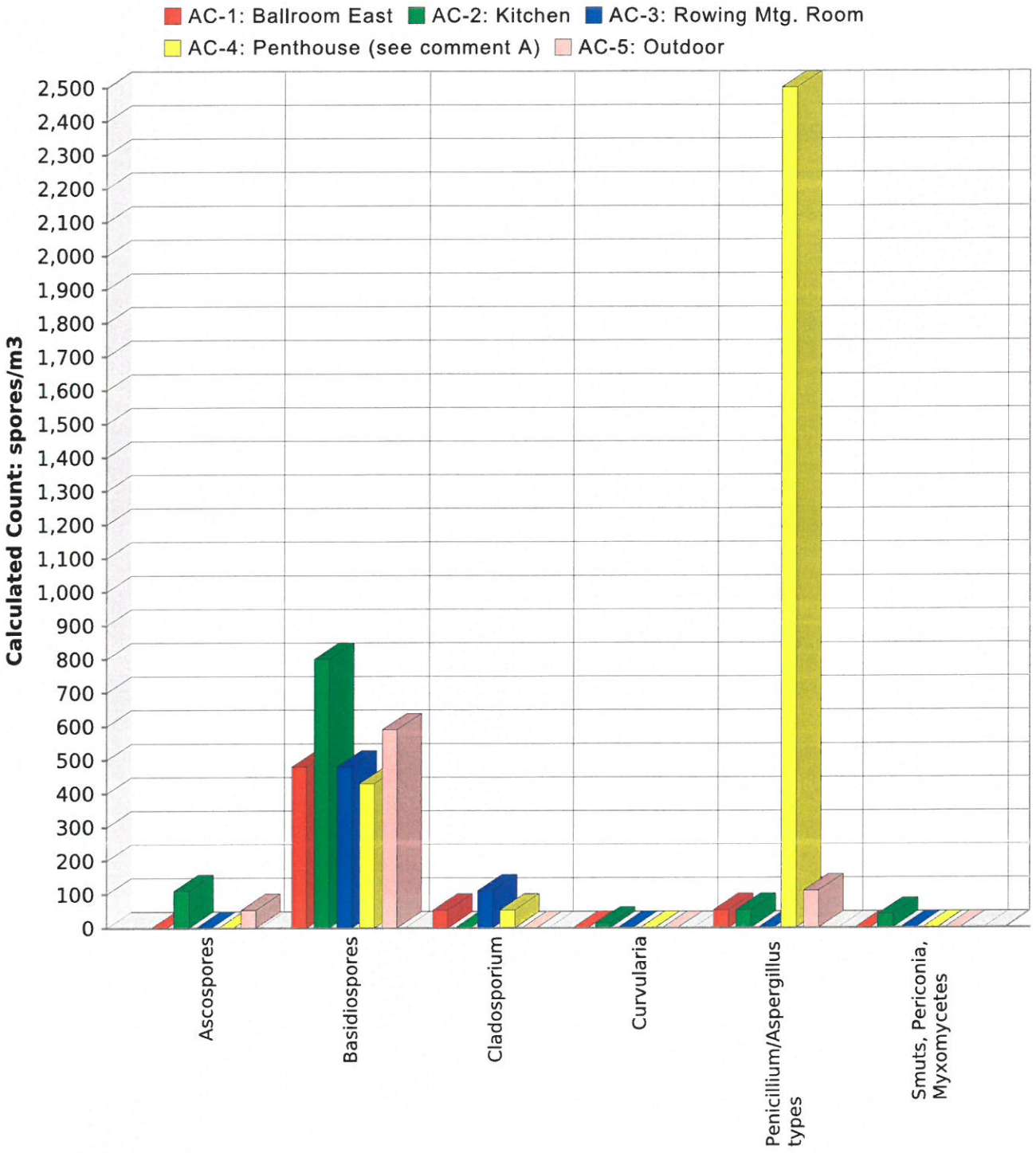
SPORE TRAP REPORT: NON-VIABLE METHODOLOGY



Comments: A) 41 of the raw count *Penicillium/Aspergillus* type spores were present as a single clump.

Note: Graphical output may understate the importance of certain "marker" genera.

CHAIN OF CUSTODY

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San Bruno, CA: 1150 Bayhill Drive, #100, San Bruno, CA 94066 * **(866) 888-6653**

003512408

Weather		Fog	Rain	Snow	Wind	Clear
Level	None	☑	☑	☑	☐	✓
	Light	☐	☐	☐	☐	
	Moderate	☐	☐	☐	☐	
	Heavy	☐	☐	☐	☑	

CONTACT INFORMATION

Company:	ACES, Inc.	Address:	PO Box 3229, Augusta, GA 30914
Contact:	Steve Connor	Special Instructions:	
Phone:	706-262-2000		

PROJECT INFORMATION

Project ID:	2006-113-002		
Project Description:	Boathouse		
Project Zip Code:	30901	Sampling Date & Time:	1-19-24/1010
PO Number:		Sampled By:	S. Connor

TURN AROUND TIME CODES (TAT)

STD – Standard (DEFAULT)
ND – Next Business Day
SD – Same Business Day Rush
WH – Weekend / Holiday

Rushes received after 2 pm or on weekends, will be considered received the next business day. Please alert us in advance of weekend analysis needs.

Sample ID	Description	Sample Type (Below)	TAT (Above)	Total Volume / Area (as applicable)	Notes (Time of day, Temp, RH, etc.)
AC-1	Ballroom East	ST	STD	75L	1010, 58.7, 35.2
AC-2	Kitchen	ST	STD	75L	1017, 57.9, 37.6
AC-3	Rowing Mtg. Room	ST	STD	75L	1027, 53.2, 44.1
AC-4	Penthouse	ST	STD	75L	1038, 73.0, 41.1
AC-5	Outdoor	ST	STD	75L	1055, 51.7, 54.8

REQUESTED SE[RVICES]

(Use checkboxes [...])

Requested service	AC-1	AC-2	AC-3	AC-4	AC-5
Non-Culturable — Spore Trap					
Fungi – Spore Trap Analysis	☑	☑	☑	☑	☑
Spore Trap Analysis – Other particles	☐	☐	☐	☐	☐
Non-Culturable — Tape Swab Bulk					
Direct Microscopic Exam (Qualitative)	☐	☐	☐	☐	☐
Quantitative Spore Count Direct Exam	☐	☐	☐	☐	☐
Culturable — BioCassette™, Andersen, SAS, Swab, Water, Bulk, Dust, Soil, Contact Plates					
1-Media Surface Fungi (Genus ID + *Asp.* spp.)	☐	☐	☐	☐	☐
2-Media Surface Fungi (Genus ID + *Asp.* spp.)	☐	☐	☐	☐	☐
3-Media Surface Fungi (Genus ID + *Asp.* spp.)	☐	☐	☐	☐	☐
Culturable Air Fungi (Genus ID + *Asp.* spp.)	☐	☐	☐	☐	☐
Gram Stain & Counts (Culturable Air & Surface Bacteria)	☐	☐	☐	☐	☐
Legionella culture	☐	☐	☐	☐	☐
Total Coliform, *E. coli (Presence/Absence)*	☐	☐	☐	☐	☐
Membrane Filtration (specify organism):	☐	☐	☐	☐	☐
MPN Bacteria (specify organism):	☐	☐	☐	☐	☐
QuantiTray – Sewage Screen	☐	☐	☐	☐	☐
Other Requests					
Asbestos Analysis – PCM Airborne Fiber Count (NIOSH 7400)	☐	☐	☐	☐	☐
Asbestos Analysis – PLM (EPA method 600/R-93-116)	☐	☐	☐	☐	☐
PCR (specify test):	☐	☐	☐	☐	☐
Specify Service	☐	☐	☐	☐	☐

SAMPLE TYPE CODES			
BC – BioCassette ™	ST – Spore Trap: Zefon, Allergenco, Burkard ...	T – Tape	D – Dust
A1S – Anderson		SW – Swab	SO – Soil
SAS – Surface Air Sampler	P – Potable Water	B – Bulk	
CP – Contact Plate	NP – Non-Potable Water	O – Other:	

RELINQUISHED BY	DATE & TIME	RECEIVED BY	DATE & TIME
[signature]	1-19-24/1215	[signature]	1/22/24 8:47AM

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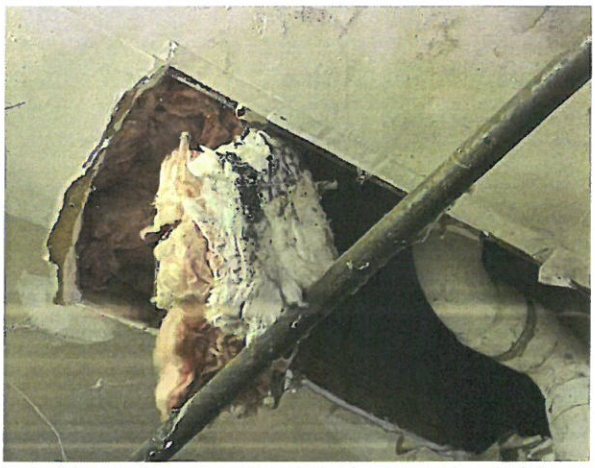
1, Basement, Rowing Club Facility

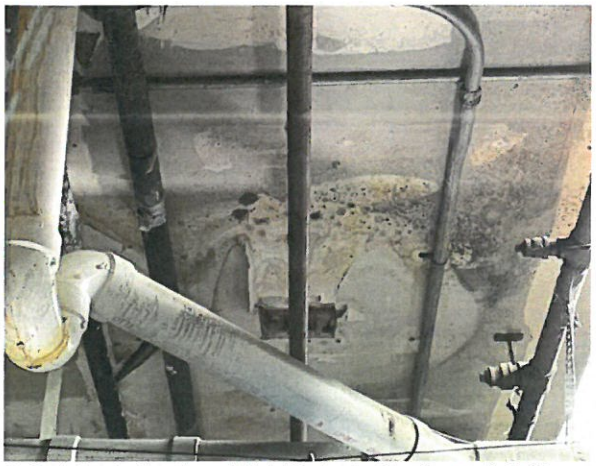


2, Staining Indicative of Mold Colonization on Insulation

3, Evidence of Water Intrusion and Damage to Drywall

4, Staining Indicative of Mold Colonization Drywall Ceiling

5, Staining Indicative of Mold Colonization on Ceiling Beam



6, Staining Indicative of Mold Colonization Drywall Ceiling