ALTERNATIVE CONSTRUCTION & ENVIRONMENTAL SOLUTIONS, INC.

2247 Wrightsboro Road
Augusta, GA 30904

Mailing Address: PO Box 3229, Augusta, GA 30914-3229
Telephone: 706-262-2000 • Facsimile: 706-262-3299 • www.aces-usa.com

October 12, 2023

Client: Kirsten Alyward

Project: Indoor Air Quality Inspection and Spore Count Analysis
Boathouse Community Center
101 Riverfront Drive
Augusta, GA 30901
Report Number: 206-101-426

1.0 SCOPE

At the request of the client, a representative of Alternative Construction & Environmental Solutions, Inc., (ACES) conducted an Indoor Air Quality visual inspection, mold sampling and analysis of the Boathouse Community Center. The area tested was limited only to that area specified by the client and was not conducted in every space of the building.

2.0 SITE/VISUAL INSPECTION

A representative of ACES conducted a visual site inspection of the above-referenced project area on October 6, 2023. This was performed to determine the current condition of the surfaces and to identify any discoloration or staining at the time of this inspection. The boat storage and workout areas were not sampled due to the space being unconditioned. Some visible discoloration was observed on the ceiling throughout those areas. There was no visible evidence of staining indicative of mold colonization found on surfaces in the other areas in the club room, ball room and penthouse. No destructive or invasive inspecting was performed. No plants or air fresheners were observed.

3.0 DISCUSSION

Sampling for mold is typically to aid in establishing 1) whether mold spores are present in an area that was designed to prevent mold from entering or forming, 2) if there is an inside source of spores that could lead to increased occupants' exposure, or 3) if there is moisture and the presence of fungi that could be an indication of possible structural damage now or sometime in the future.

- ***Air Samples:***
  Air samples are divided into two different methods, culturable and non-culturable (non-viable or "spore trap"). Both samples are acquired using an air pump attached to a media.

  A. BioCassettes® with a Malt Extract Agar (MEA) are used for the determination of culturable (viable) fungi present in the air. Air sample duration is from one to five minutes at 28.3 liters per minute based on visual evaluation of the areas level of suspected contamination.

  B. Non-culturable samples are acquired using Air-O-Cell® spore trap samplers. This is a particulate sampling cassette for the rapid collection and analysis of a wide range of mold spores. This type of sampling does not allow differentiation between viable and non-viable mold spores. Air sample duration is determined by an expected level of contamination chart with sample times of 0.5 – 10 minutes at 15 liters per minute.

  "Air Sampling is limited, and negative results do not document the absence of mold exposure. For example, mold may be growing in carpets or on walls and wallpapers, yet not be airborne at the time of sampling. Where there are other indications, such as moisture noted where it should not be, further investigation for hidden sources is indicated." (ref. 1)

- ***Surface Samples:***
  Surface samples are typically taken by tape lift imprint, by swabbing the surface of suspected mold growth with a culturette swab, or by destructive sampling of the suspected mold growth materials. These samples are submitted to the laboratory for analysis for microbial contamination. The laboratory reports the findings by direct microscopic examination to identify the types of mold growth.

"There is substantial natural variability in the amount of mold in air. Understandably, the EPA and other government agencies have not set numeric standards for indoor concentrations of mold or mold spores." (ref. 1)

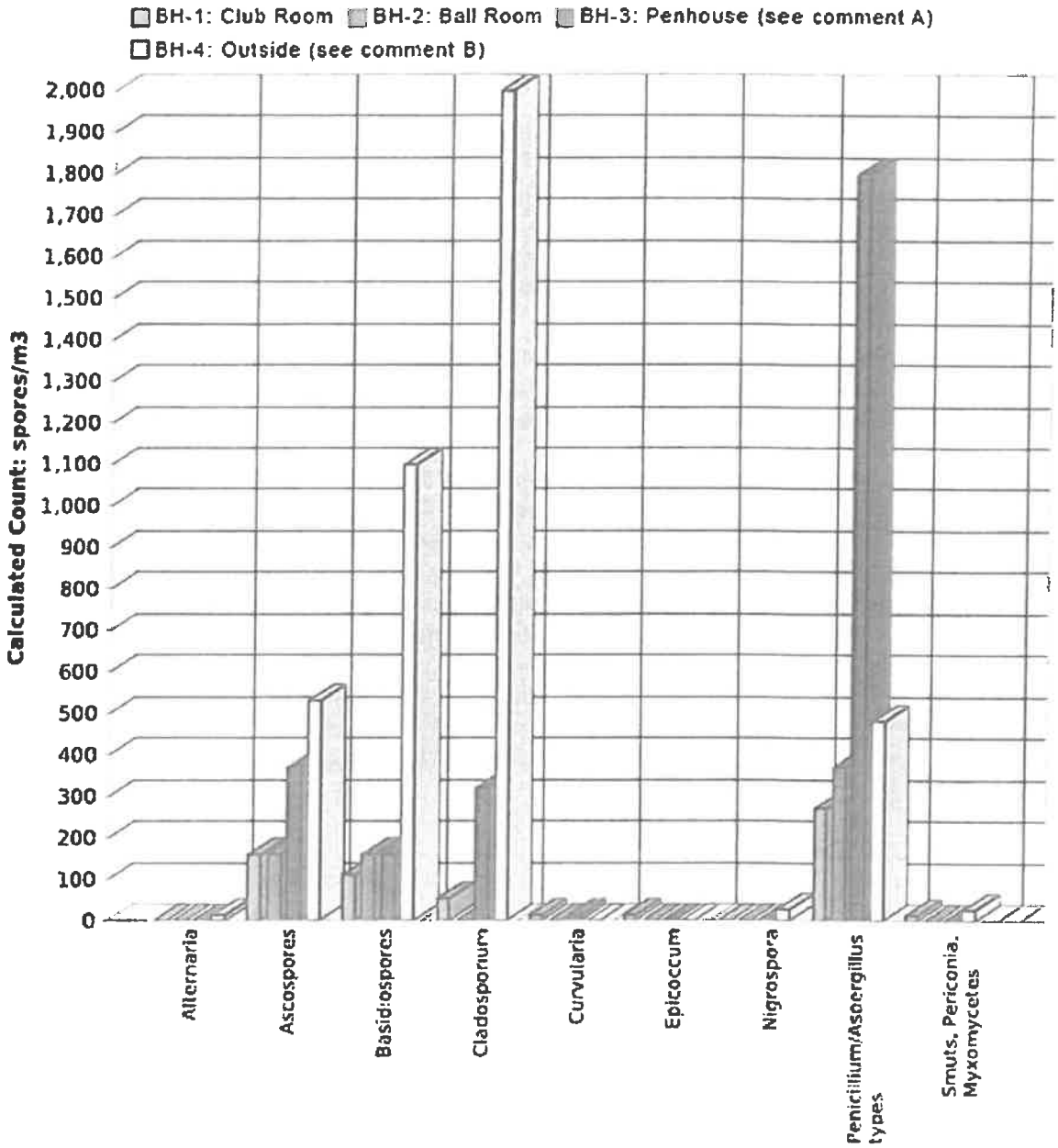
4.0 SAMPLING AND ANALYSIS

Sampling was accomplished utilizing a Zefon Bio-Pump with Air-O-Cell® cassettes. Four Air-O-Cell® samples were taken from various areas to include both inside and outside samples. The number of samples taken during this investigation was specified by the client.

Samples collected were assigned a unique sample ID number and placed in a sealed container. Samples were sent to EMLab P&K (Environmental Microbiology Laboratory, Inc.), an American Industrial Hygiene Association (AIHA) Environmental Microbiology-accredited laboratory (#178699).

Table I presents a graphical depiction of the spore counts obtained from this project. Sample BH-1 is depicted in red for the inside sample obtained in the Club Room. Sample BH-2 is depicted in green for the inside sample obtained from Ball Room. Sample BH-3 is depicted in blue for the inside sample obtained in the Penthouse. Sample BH-4 is depicted in for the outside sample.

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY



Comments: Table 1

The Bioaerosol Committee of the American Conference of Governmental Industrial Hygienists (ACGIH) states that outdoor airborne fungi concentration "routinely exceeds 1,000 CFU/m^3 and may average near 10,000 CFU/m^3 in summer months." No occupational exposure limit for bioaerosols has been promulgated by the Occupational Safety and Health Administration (OSHA). (ref. 3)

Additionally, temperature, relative humidity, carbon dioxide and carbon monoxide readings were taken using the Gray Wolf Indoor Air Quality Probe MP Surveyor Pro with serial number 78014.

Location	Time	Temp. (°F)	Relative Humidity (%)	Carbon Dioxide (ppm)	Carbon Monoxide (ppm)
Club Room	0917	76.3	**60.2**	304	2.6
Ball Room	0924	77.1	**59.4**	239	1.5
Penthouse	0929	76.6	**58.7**	275	1.0
Outside	0936	72.4	70.7	226	1.2

The American Society of Heating, Refrigerating, and Air-Conditioning Engineers (ASHRAE) standard "Ventilation for Acceptable Air Quality", establishes 1000 parts per million (ppm) Carbon Dioxide as a level above which ventilation may be inadequate. (ref. 2)

Elevated Carbon Monoxide levels in indoor air indicate existence of a combustion source exhaust that is not effectively being ventilated out of the building. The standard of 25 ppm established by the American Conference of Governmental Industrial Hygienist (ACGIH) is widely accepted as a limit for defining hazardous carbon monoxide levels. (ref. 3) The ASHRAE ventilation standard has established 9 ppm or greater of carbon monoxide within an occupied space as being a level of concern. (ref. 2) The Environmental Protection Agency defines 9 ppm as a limit for acceptable air quality.

"The Relative Humidity (RH) of the indoor air and ventilation system should be below 60 percent. Ideally, RH should be kept between 30 and 50 percent because, at a RH of 50 percent or more, hydroscopic dust will absorb water that may allow the growth of fungi and house dust mites on indoor surfaces." (ref. 1)

5.0 ATTACHMENTS

Attachment I Lab Results

6.0 CONCLUSIONS

The boat storage areas and the workout areas on the bottom floor were not sampled. This space is and has been an unconditioned space, relatively open to the outside air. The discoloration in the boat storage areas and the workout areas appeared to be indicative of the unconditioned space and years of buildup on the gypsum board ceiling and does not appear to be the result of a water event.

No other discoloration was observed in the areas sampled. However, no destructive or invasive inspecting was performed.

A single Epicoccum spore was identified in the club room sample but was not in the outside sample. The Penicillium/Aspergillus count in the penthouse was higher than the outdoor sample. All other spores found in the sampled areas were at or below the outside levels. This data suggests that there could be a potential indoor source in the penthouse.

The following is a list of the spores found inside the project area with a brief description of each.

Ascospores are found everywhere in nature, particularly in Saprophytes and plant pathogens. The spores are predominantly forcibly discharged during periods of high humidity or rain.

Basidiospores is a common spore found outdoors on decaying wood and on plants. In the indoor environment, it is one of the fungi responsible for causing white or brown wood rot if allowed to form large colonies and can sometimes be recognized by the presence of mushrooms. These spores can grow to destroy the structural wood of buildings.

Cladosporium is typically found on leaves and decaying plants in the outdoor environment. In the indoor environment, it can be found in insulation and on window panes that have colder surfaces.

Curvularia in the indoor environment is typically from plant debris, soil, and facultative plant pathogens of tropical or subtropical plants. More commonly found in tropical, subtropical regions.

Epicoccum in the indoor environment is typically from plant debris and soil.

Penicillium/Aspergillus in the indoor environment is typically from water-damaged materials.

Smuts, Periconia, Myxomycetes are not typically found indoors. They are parasitic plant pathogens that require a living host for the completion of their life cycle.

According to the MoldRANGE™ Extended Outdoor Comparison Chart, the Penicillium/Aspergillus were higher than the very high range outdoor data for the month of October in the State of Geogia. All other spore counts recorded in this survey were below the typical very low outdoor data comparison for the month of October in the State of Georgia.

No occupational exposure limit for bioaerosols has been promulgated by the Occupational Safety and Health Administration (OSHA). "If fungal concentrations indoors are consistently higher than those outdoors, then indoor sources are indicated." (ref. 3)

Most spores found during this sampling are primarily found in the soil, leaves, grass, weeds, and other types of vegetation and can be easily transmitted into the building by people moving in and out.

The Relative Humidity levels found inside during this inspection were above 50%. A Relative Humidity of 50% or greater can increase the chances of fungi growth. The function of the HVAC system should be evaluated to ensure that levels are kept within the ideal range of 30-50%.

If any materials are found in the future with visual mold growth or damaged appearance, they should be cleaned or removed/discarded as per EPA guidelines published in "Mold Remediation in Schools and Commercial Buildings" (EPA 402-K-01-001). (ref. 4)

It should be noted that microbiological growth can and probably will reoccur if any source of moisture is not corrected and maintained as appropriate. Many spores are naturally occurring and will grow again with proper moisture, temperature and food source.

This report relates only for this time and conditions present during our investigation. Facilities are constantly under influence from several external factors, activities, environmental conditions and pollutants that are subject to change. If any new or conflicting information becomes available at a later date, please advise ACES and any appropriate revisions and/or comments will be made. This report was prepared for the client and should not be reproduced, except in whole and only with the written approval of ACES.

ALTERNATIVE CONSTRUCTION & ENVIRONMENTAL SOLUTIONS, INC.

Cliff Hampton
Manager
Industrial Hygiene, Safety & Health Dept.

Austin Metcalf
Project Manager

Attachment

References:

1. University of Connecticut Health Center Division of Occupational and Environmental Medicine, Center for Indoor Environments and Health, *Guidance for Clinicians on the Recognition and Management of Health Effects Related to Mold Exposure and Moisture Indoors, (September 30, 2004)*

2. American Society of Heating, Refrigerating and Air-Conditioning Engineers, Inc. (*ANSI/ASHRAE 62-1989), Ventilation for Acceptable Air Quality*, ASHRAE, Atlanta, GA 1992.

3. American Conference of Governmental Industrial Hygienists (ACGIH), *Bioaerosols: Assessment and Control* (Edited by J. Marcher), Cincinnati, OH 1999.

4. Environmental Protection Agency (EPA), *Mold Remediation in Schools and Commercial Buildings* (EPA 402-K-01-001, March 2001)

eurofins

Report for:

Austin Metcalf
Alternative Construction and Environmental Solutions
P.O. Box 3229
Augusta, GA 30914

Regarding: Eurofins EPK Built Environment Testing, LLC
Project: 206-101-426; 101 Riverfront Dr. Augusta, GA 30901
EML ID: 3412693

Approved by:

Business Unit Manager
Balu Krishnan

Dates of Analysis:
Spore trap analysis: 10-09-2023

Service SOPs: Spore trap analysis (EB-MY-S-1038)
AIHA-LAP, LLC accredited service, Lab ID #221504

All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank correction of results is not applied. The results relate only to the samples as received and tested. Information supplied by the client which can affect the validity of results: sample air volume.

Eurofins EPK Built Environment Testing, LLC
6215 Regency Parkway, Suite 900, Norcross, GA 30071
(866) 871-1984 www.eurofinsus.com/Built

Client: Alternative Construction and Environmental Solutions
C/O: Austin Metcalf
Re: 206-101-426; 101 Riverfront Dr. Augusta, GA 30901

Date of Sampling: 10-06-2023
Date of Receipt: 10-09-2023
Date of Report: 10-10-2023

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	BH-1: Club Room		BH-2: Ball Room		BH-3: Penhouse		BH-4: Outside	
Comments (see below)	None		None		A		B	
Lab ID-Version‡:	16602030-1		16602031-1		16602032-1		16602033-1	
Analysis Date:	10/09/2023		10/09/2023		10/09/2023		10/09/2023	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria							1	13
Arthrinium								
Ascospores	3	160	3	160	7	370	10	530
Aureobasidium								
Basidiospores	2	110	3	160	3	160	20	1,100
Bipolaris/Drechslera group								
Botrytis								
Chaetomium								
Cladosporium	1	53			6	320	117	2,000
Curvularia	1	13			1	13		
Epicoccum	1	13						
Fusarium								
Myrothecium								
Nigrospora							2	27
Other colorless								
Penicillium/Aspergillus types†	5	270	7	370	45	1,800	9	480
Pithomyces								
Rusts								
Smuts, Periconia, Myxomycetes	1	13					2	27
Stachybotrys								
Stemphylium								
Torula								
Ulocladium								
Zygomycetes								
Background debris (1-4+)††	2+		2+		3+		3+	
Sample volume (liters)	75		75		75		75	
§ TOTAL SPORES/m3		630		690		2,600		4,100

Comments: A) 16 of the raw count *Penicillium/Aspergillus* type spores were present as a single clump. B) 107 of the raw count *Cladosporium* spores were present as a single clump.

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.
† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium, Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.
††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m^3 divided by the raw count, expressed in spores/m^3, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.
‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".
§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.

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6215 Regency Parkway, Suite 900, Norcross, GA 30071
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Client: Alternative Construction and Environmental Solutions
C/O: Austin Metcalf
Re: 206-101-426; 101 Riverfront Dr. Augusta, GA 30901

Date of Sampling: 10-06-2023
Date of Receipt: 10-09-2023
Date of Report: 10-10-2023

MoldRANGE™, Local Climate; Extended Outdoor Comparison

Outdoor Location: BH-4, Outside

Fungi Identified	Outdoor data	Typical Outdoor Data for: October in Southeast† EMLab Regional Climate code¹ A Annual Temp, A Elev., B Rain, A Temp. Range (n‡=327)						Typical Outdoor Data for: The entire year in Southeast† EMLab Regional Climate code¹ A Annual Temp, A Elev., B Rain, A Temp. Range (n‡=3146)					
Project zip code 30901	spores/m3	very low	low	med	high	very high	freq %	very low	low	med	high	very high	freq %
Generally able to grow indoors*													
Alternaria	13	13	13	27	67	110	48	13	13	27	80	120	40
Bipolaris/Drechslera group	-	13	13	13	40	110	33	13	13	13	53	94	21
Chaetomium	-	-	-	-	-	-	4	13	13	13	40	85	5
Cladosporium	2,000	160	270	910	2,400	4,300	98	53	130	590	1,800	3,100	91
Curvularia	-	13	13	27	67	160	52	13	13	27	80	150	31
Epicoccum	-	13	13	27	40	53	34	13	13	27	53	99	32
Nigrospora	27	13	13	21	53	76	36	7	13	13	40	53	20
Penicillium/Aspergillus types	480	53	110	290	1,100	1,500	80	40	67	210	690	1,200	76
Stachybotrys	-	-	-	-	-	-	2	9	13	27	77	180	1
Torula	-	13	13	13	40	53	17	8	13	13	47	80	12
Seldom found growing indoors**													
Ascospores	530	110	160	480	1,400	2,300	97	53	110	520	1,900	3,500	90
Basidiospores	1,100	530	950	4,100	15,000	26,000	99	160	400	2,300	11,000	22,000	98
Rusts	-	13	13	27	80	120	24	13	13	27	67	160	20
Smuts, Periconia, Myxomycetes	27	27	33	93	230	290	86	13	20	53	150	240	71
§ TOTAL SPORES/m3	4,100												

¹EMLab Regional Climate codes are a climate classification scheme for regional geographic areas containing multiple states. The MoldRANGE™ Local Climate report uses the sampling location zip code to identify the EMLab Regional Climate code in that area. Using information available from the NOAA weather database, the EMLab Regional Climate code sharpens the precision of the MoldRANGE™reporting system, providing more reliable estimates of the range and average concentrations of the different airborne fungal spore types for each region. Additional information on the EMLab Regional Climate code system can be found on the last page of this report.

†The Typical Outdoor Data represents the typical outdoor spore levels across the region's group of states for the time period and EMLab Regional Climate code indicated. The last column represents the frequency of occurrence. The very low, low, med. high, and very high values represent the 10, 20, 50, 80, and 90 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 20% of the time it is present in levels above the detection limit and below 53 spores/m3. These values are updated periodically and if not enough data is available to make a statistically meaningful assessment, it is indicated with a dash.

‡ n is the sample size used to calculate the MoldRANGE™ Local Climate data summarized in the table.

* The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

** These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.

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Date of Sampling: 10-06-2023
Date of Receipt: 10-09-2023
Date of Report: 10-10-2023

Understanding EMLab Regional Climate Codes

Outdoor airborne spore concentrations are strongly influenced by climate and weather patterns, often resulting in pronounced seasonal and diurnal cycles (Burge 1995). The seasonal climatic changes directly affect the growth cycle of plants, thereby influencing fungal growth, spore maturation, and release cycles. By evaluating outdoor spore concentrations across similar climatic zones rather than for the state as a whole, it is possible to provide a more representative estimate of typical outdoor spore levels and frequency of occurrence for different airborne fungal spore types in a given area.

The EMLab Regional Climate code system is a novel classification system that uses data from the NOAA - National Oceanic and Atmospheric Administration database to define unique climate zones. The following climate variables, for each regional zip code, are obtained from NOAA and assigned a letter code of A (above the regional average for that variable) or B (below the regional average for that variable):

1. Annual High Temperature
2. Elevation
3. Rainfall/Precipitation
4. Monthly Temperature Range

The result is a 4-character code assigned to each statewide zip code, referred to as the Regional Climate Code. Below are some examples of decoded Regional Climate Codes:

AAAA = Above avg. Annual High Temperature, Above avg. Elevation, Above avg. Rainfall/Precipitation, Above avg. Monthly Temperature Range
AABB = Above avg. Annual High Temperature, Above avg. Elevation, Below avg. Rainfall/Precipitation, Below avg. Monthly Temperature Range
BBAA = Below avg. Annual High Temperature, Below avg. Elevation, Above avg. Rainfall/Precipitation, Above avg. Monthly Temperature Range

The actual outdoor air sample data from matching regional climate codes in each group of states are then compiled in a manner relating typical spore concentrations and frequency of occurrence.

The data presented in this report is from the Southeast Region which includes the states of: AL, FL, GA, NC, SC, and VA

The NOAA regional climate variables were selected by mapping data points from a subset of approximately 145,000 weather and geographic database entries to over 80,000 outdoor spore trap samples with known zip codes and assessing them using orthogonal array experimental design techniques. The results were then compared to the typical ranges of spore types found when grouping zip codes using the Koppen-Geiger climatic classification system; a commonly used climatic system that provides an objective numerical definition in terms of climatic elements such as temperature, rainfall, and other seasonal characteristics . The EMLab Regional Climate codes showed improved granularity and refinement of the zip code groupings, implying a better representation of the expected range of spore types to be found within an individual zip code.

The values on this report were calculated by obtaining the four variables listed above from the over 585 million data points of weather and geographic information available in the NOAA database, and determining the frequencies and percentile values of spore types by utilizing over 180,000 Eurofins EMLab P&K outdoor spore trap samples with known zip codes.

This report groups regional zip codes in relation to these EMLab Regional Climate codes and summarizes MoldRANGE™ data by month and year within each EMLab Regional Climate code.

References:

Burge, Harriet, A. Bioaerosols: Boca Raton: Lewis Publishers, pp. 163-171, 1995.

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Date of Sampling: 10-06-2023
Date of Receipt: 10-09-2023
Date of Report: 10-10-2023

About Your New MoldRANGE Local Climate Report:

Why am I receiving this report?

The MoldRANGE Local Climate Report is a new supplemental report that contains more specific outdoor reference information for your area. We want you to see this new report so you can decide if it would be a useful tool for your investigations. Eurofins EMLab P&K is the only laboratory that equips you with scientifically-developed and statistically accurate tools like this new report for your mold investigations.

What does this report tell me?

Since the climate and outside spore distribution across even a single state may vary depending upon your location, this report breaks data from states with similar climates into smaller groupings (called EMLab Regional Climate codes) based upon climate data from the NOAA National Climactic Data Center. The spore type data comes from Eurofins EMLab P&K's unparalleled database of over 350,000 spore trap samples. We evaluated each grouping and displayed the typical frequency and counts of a given spore type within that EMLab Regional Climate code. This gives you a point of reference for the number and types of spores usually present in the outdoor air in your area across the month of sampling and the entire year.

What is the cost of this new report?

There is none. MoldRANGE Local Climate is offered as free supplement to Eurofins EMLab P&K clients.

What if I am happy with my existing report preferences and do not wish to receive this report in the future?

Simply contact your Project Manager or email us at EBET.customerservice@et.eurofinsus.com and let us know. We can reset your preferences so that you continue to receive your typical report formats instead.

10-10-2023: 206-101-426

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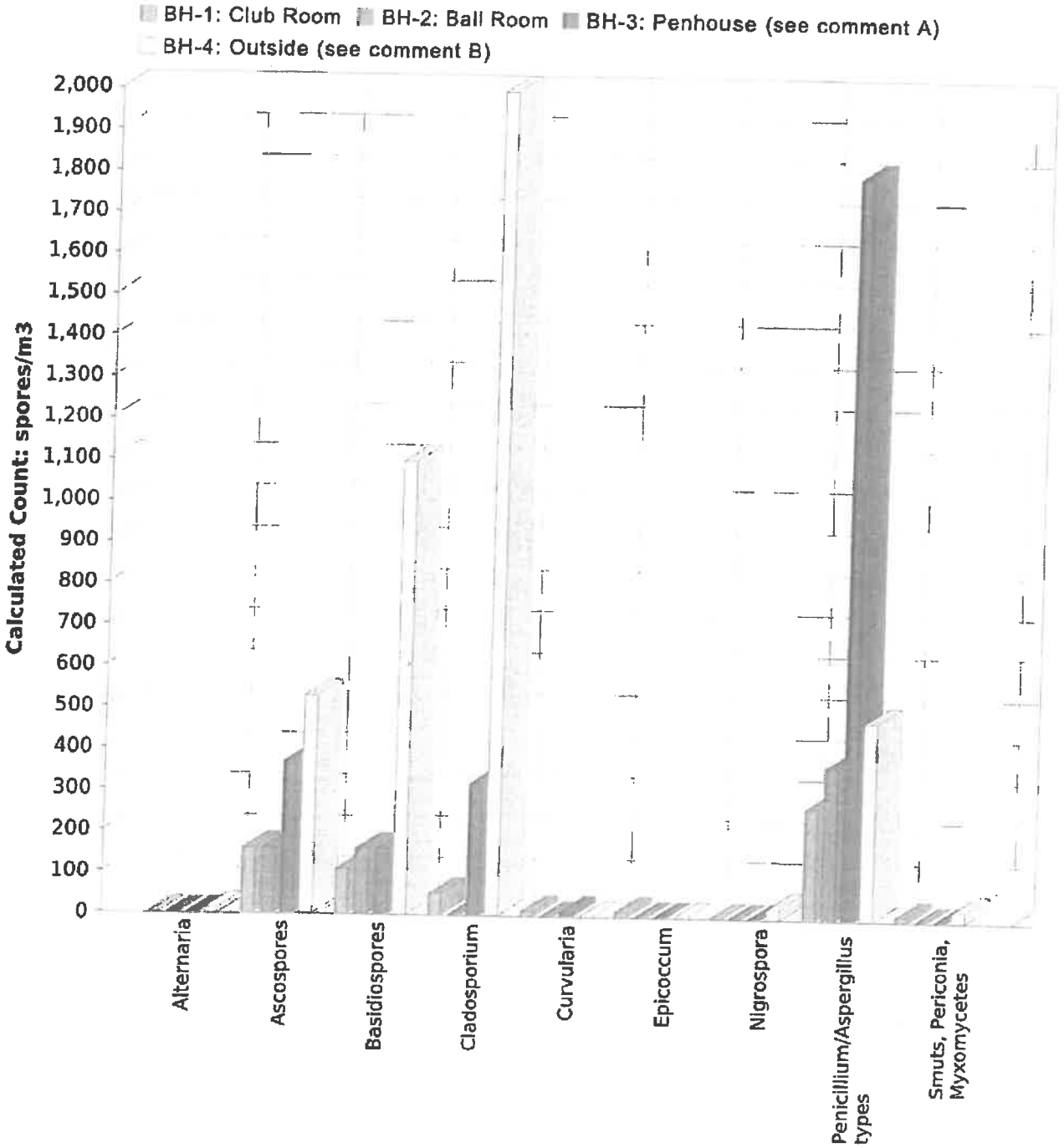
SPORE TRAP REPORT: NON-VIABLE METHODOLOGY



Comments: A) 16 of the raw count *Penicillium/Aspergillus* type spores were present as a single clump. B) 107 of the raw count *Cladosporium* spores were present as a single clump.

Note: Graphical output may understate the importance of certain "marker" genera.

CHAIN OF CUSTODY — eurofins

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East: (866) 871-1984
Central: (800) 651-4802
West: (866) 888-6653

Weather	Fog	Rain	Snow	Wind	Clear
Level: None	☐	☐	☐	☐	☐
Light	☐	☐	☐	☐	
Moderate	☐	☐	☐	☐	
Heavy	☐	☐	☐	☐	

Ptly Cloudy

FUNGAL A

003412693

CONTACT INFORMATION

Company: ACGS

Contact: Austin Metcalf

Phone: 706-762-2000

Address: 2247 Wrightsboro Rd. [illegible], GA

Special Instructions: Send Results to results@[illegible].com

PROJECT INFORMATION

Project ID: 206-101-4126

Project Description: 101 Riverfront Dr. Augusta, GA 30901

Project Zip Code: 30901

Sampling Date & Time: 10/6/23 0915

PO Number:

Sampled By: Austin Metcalf

TURN AROUND TIME CODES (TAT)

STD – Standard (DEFAULT)

ND – Next Business Day

SD – Same Business Day Rush

WH – Weekend / Holiday

Rushes received after 2pm or on weekends, will be considered received the next business day. Please alert us in advance of weekend analysis needs.

REQUESTED SERVICES
(Check boxes below)

Non-Culturable: Spore Trap; Tape Swab Bulk

Culturable: BioCassette™, Andersen, SAS, Swab, Water, Bulk, Dust, Soil, Contact Plates

Sample ID	Description	Sample Type (Below)	TAT (Above)	Total Volume / Area (as applicable)	Notes (Time of day, Temp, RH, etc.)	Spore Trap Analysis
BH-1	Club Room	ST	STD	75 liters	0911 16.5°C 60.2%	☒
BH-2	Ball Room	ST	STD	75 liters	0924 22.5°C [illegible]	☒
BH-3	[illegible]	ST	STD	75 liters	0929 16.6°C [illegible]	☒
BH-4	Outside	ST	STD	75 liters	0936 17.5°C [illegible]	☒

Other requested services (not checked): Spore Trap Analysis – Other particles; Direct Microscopic Exam (Qualitative); Quantitative Spore Count Direct Exam; Dust Characterization; Air Fungi (Genus ID + Asp. speciation); Air Fungi - Full speciation Pen. & Clad. genus only; Air Fungi - Full speciation; 1-Media Surface Fungi (Genus ID + Asp. speciation); 1-Media Surface Fungi - Full spec. Pen/Clad genus only; 1-Media Surface Fungi - Full speciation; 2-Media Surface Fungi (Genus ID + Asp. speciation); 2-Media Surface Fungi - Full spec. Pen/Clad genus only; 2-Media Surface Fungi - Full speciation; 3-Media Surface Fungi (Genus ID + Asp. speciation); 3-Media Surface Fungi - Full spec. Pen/Clad genus only; 3-Media Surface Fungi - Full speciation; Other

SAMPLE TYPE CODES

BC – BioCassette™	CP – Contact Plate	T – Tape	D – Dust
A1S – Andersen	ST – Spore Trap Zefon, Allergenco, Burkard …	SW – Swab	W – Water
SAS – Surface Air Sampler		B – Bulk	SO – Soil
O – Other			

RELINQUISHED BY	DATE & TIME	RECEIVED BY	DATE & TIME
A. C. Metcalf	10-6-23 1030	[illegible]	10/9/23 11:18

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